

RAPID COMMUNICATIONS

CONVERSION OF THE HUMAN 5-HT_{1D}β SEROTONIN RECEPTOR TO THE RAT 5-HT_{1B} LIGAND-BINDING PHENOTYPE BY THR³⁵⁵ASN SITE DIRECTED MUTAGENESIS

Mark A. Metcalf*, Robert W. McGuffin*†‡ and Mark W. Hamblin*†§

*Geriatrics Research, Education and Clinical Center, Seattle Veterans Affairs Medical Center, and the

†Department of Psychiatry and Behavioral Sciences, University of Washington School of Medicine,
Seattle, WA 98108, U.S.A.

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ABSTRACT — The human 5-HT_{1D}β serotonin receptor and its rat homolog (also called the 5-HT_{1B} receptor) share 93% amino acid identity, yet display markedly different pharmacological specificities. Comparison of deduced amino acid sequences among these and other recently cloned receptors suggested that this phenotypic difference might be attributable to a single human threonine³⁵⁵/rat asparagine³⁵¹ amino acid difference in the putative seventh membrane spanning regions. We now report that Thr³⁵⁵Asn mutagenesis of the human 5-HT_{1D}β receptor alters the binding characteristics of the recombinant receptor in [³H]5-HT binding assays to a profile very similar to that of the rat 5-HT_{1B} binding site. These results confirm that this single amino acid difference is responsible for the majority of the known pharmacological discrepancies between human and rat observed for 5-HT_{1D}β (5-HT_{1B}) receptors.

Recent months have seen an explosion in the number of receptors for serotonin (5-hydroxytryptamine, 5-HT) that have been cloned. We have cloned two human 5-HT_{1D} receptors, members of the seven α-helix G protein linked receptor family, as well as their rat homologs [1-3], and similar results have been reported independently by others [4-10]. Pharmacological characterization of these gene products has shown that one such 5-HT_{1D} receptor subtype, also referred to as the 5-HT_{1D}α receptor, shows nearly identical pharmacological profiles in human and rat [3]. The other currently known 5-HT_{1D} receptor shows a classical 5-HT_{1D}-like receptor pharmacological profile in human, but a strikingly different, 5-HT_{1B}-like profile in rat [3,8,9] and mouse [10]. The 5-HT_{1B} phenotype is distinguished from the classic 5-HT_{1D} profile chiefly by its much higher affinity for pindolol and its derivatives and somewhat higher affinity for the indole RU 24969. Consequently, this second human 5-HT_{1D} receptor has been referred to as either the 5-HT_{1D}β receptor [6,7] or the human 5-HT_{1B} receptor [2,4].

The unusual degree of species-specific pharmacological difference for the 5-HT_{1D}β receptor was not entirely unexpected, based on observations from animal tissues and cells (reviewed in [11]). Nearly identical 5-HT_{1B}-like phenotypes have been observed in some rodents and the evolutionarily quite distantly related American Opossum, but no receptors with intermediate 5-HT_{1B}/5-HT_{1D}-hybrid phenotypes have been found in other mammalian species. These observations suggested that the species-specific differences in the 5-HT_{1B}/5-HT_{1D} receptor family might be due to a single amino acid difference, or perhaps at most a very few. Just where such a difference might lie was suggested by the work of Kobilka and colleagues on other G protein coupled receptors [12]. In a study of chimeric adrenergic receptors, high affinity for [¹²⁵I]cyanopindolol, a key characteristic of β-adrenergic receptors as well as 5-HT_{1B} receptors, was found to be conferred by the predicted seventh membrane spanning region. More recent site directed mutagenesis studies have shown that a single critical amino acid position, corresponding to Thr³⁵⁵ in the human 5-HT_{1D}β receptor, greatly influences the drug-binding profile of adrenergic [13] as well as 5-HT_{1A} serotonergic receptors [14]. The presence of an asparagine in this position correlates with high pindolol affinity.

‡ Current address: Department of Psychiatry (0603), UCSD School of Medicine, La Jolla, CA 92093.

§ Corresponding author: Dr. Mark Hamblin, GRECC (182B), Seattle Veterans Affairs Medical Center, 1660 South Columbian Way, Seattle, WA 98108. Tel. (206) 764-2308; FAX (206) 764-2569.

We, as well as Adham and co-workers, noted that the cloned rat 5-HT_{1B} receptor also has a predicted asparagine residue in this critical position in the predicted seventh hydrophobic region, while its human homolog, the 5-HT_{1Dβ} receptor, has a threonine in this position [2,8], as do the human [1] and rat [3] 5-HT_{1Dα} receptors. We wished to test directly the hypothesis that this single amino acid difference provides much of the pharmacological distinction between rat and human 5-HT_{1Dβ} (5-HT_{1B}) receptors. In this paper we describe the pharmacological analysis of a Thr³⁵⁵Asn mutant of the human 5-HT_{1Dβ} receptor. We show that this mutated receptor is altered to the classic rat 5-HT_{1B}-like (high cyanopindolol-affinity) phenotype.

MATERIALS AND METHODS

Construction of wild type and Thr³⁵⁵Asn mutant human 5-HT_{1Dβ} receptor constructs for expression. A 528 bp Xma I fragment of the human 5-HT_{1Dβ} (5-HT_{1B}) receptor [2] which encoded the presumed seventh membrane spanning region was subcloned into pBluescript II KS(-) in the T3→T7 orientation. This served as a template for mutation using the polymerase chain reaction-based method of Ito and co-workers [15]. The mutant primer employed was 5'-ATAGCCCAGCCAATTGAAGAAGTCAAAG-3'. Both strands of the mutated Xma I fragment were then sequenced to ensure that no inadvertent mutations had been introduced. The mutated Xma I fragment was then excised and ligated into the adjacent 1.3 kb Hind III-Xma I fragment of the receptor gene, also in pBluescript II KS(-) in the T3→T7 orientation, to recreate the entire, now mutated open reading frame. A Xho I-Bam HI fragment containing the open reading frame was then subcloned into the eukaryotic expression vector pSVL (Pharmacia), in the proper orientation. A corresponding wild type Bam HI fragment of the unaltered gene was also subcloned into pSVL for wild type receptor expression.

Receptor expression and [³H]5-HT binding assays. Wild type or mutant expression construct DNA was used to transfect COS-7 cells (20 µg/150 mm culture plate) using the method described in Ref. 16. After 48-72 hr, the cells were harvested by scraping into Dulbecco's phosphate-buffered saline supplemented with 5 mM EDTA and assessed for [³H]5-HT (24-29 Ci/mmol, New England Nuclear) binding as previously described [3]. Sources of drugs were as described in Ref. 1. IC₅₀ values were determined by the method of Parker and Waud [17].

RESULTS AND DISCUSSION

We constructed a human 5-HT_{1Dβ} threonine³⁵⁵asparagine mutant to see if this change would explain the great interspecies pharmacological differences found for this receptor. The affinities of drugs examined for the wild type human 5-HT_{1Dβ} receptor expressed in COS-7 cells as assessed in [³H]5-HT competition binding assays (Table 1) were in good agreement with values that we previously obtained using stably transfected HeLa cells [2]. Introduction of the Thr³⁵⁵Asn mutation had no discernable effect on the affinity of 5-HT. However, the mutation increased the affinity of the receptor for (±)cyanopindolol by 180-fold and for RU 24969 by 30-fold (Table 1), placing the affinities of these two compounds in the range of what would be expected of a rat 5-HT_{1B} receptor. Although not generally regarded as a defining difference between 5-HT_{1D} and 5-HT_{1B} receptor profiles, studies of

Table 1. Effect of the Thr³⁵⁵Asn mutation on drug affinity for [³H]5-HT (2 nM) labeled 5-HT_{1Dβ} receptors expressed in COS-7 cells

Drug	Human 5-HT _{1Dβ} wild type IC ₅₀ (nM)		Human 5-HT _{1Dβ} Thr ³⁵⁵ Asn mutant IC ₅₀ (nM)		Mutation-induced shift (fold)
(±)Cyanopindolol	150	± 30	0.84	± 0.28	180
RU 24969	97	± 19	3.3	± 0.2	30
5-HT	5.5	± 2.3	5.5	± 0.1	1.0
Sumatriptan	14	± 4	190	± 30	(13)
CGS 12066B	2.1	± 0.6	30	± 9	(14)
Yohimbine	63	± 13	2,000	± 300	(31)
8-OH-DPAT	320	± 20	18,000	± 1,600	(56)

IC₅₀ values are expressed as means ± SEM for N = 3.

brain tissues in the appropriate species have generally found (\pm)-8-hydroxy-*N,N*-dipropyl-2-aminotetralin (8-OH-DPAT) to have substantially greater affinity for 5-HT_{1D} type brain binding sites [18-20] than 5-HT_{1B} sites [21]. As would be predicted by these differences, the Thr³⁵⁵Asn mutation decreased the affinity of the recombinant human 5-HT_{1D} β receptor for 8-OH-DPAT by 56-fold. Indeed, brain membrane 5-HT_{1D} binding assays have generally been performed in the presence of 100 nM 8-OH-DPAT, a practice which may decrease the apparent affinity of the human 5-HT_{1D} β receptor for this compound and minimize the apparent human versus rat 5-HT_{1D} β selectivity for 8-OH-DPAT. Interestingly, an Asn³⁸⁵Val mutation in the corresponding position of the 5-HT_{1A} receptor had little effect on the affinity of this receptor for 8-OH-DPAT [14]. Introduction of the Thr³⁵⁵Asn mutation also decreased the affinity of the 5-HT_{1D} β receptor for yohimbine. This alteration is in keeping with the observations of Suryanarayana and co-workers, who also found a decreased yohimbine affinity for a human α_2 -adrenergic receptor with the corresponding Phe⁴¹²Asn mutation [13]. Affinities for the anti-migraine drug sumatriptan and CGS 12066B were also decreased for the mutant human 5-HT_{1D} β receptor.

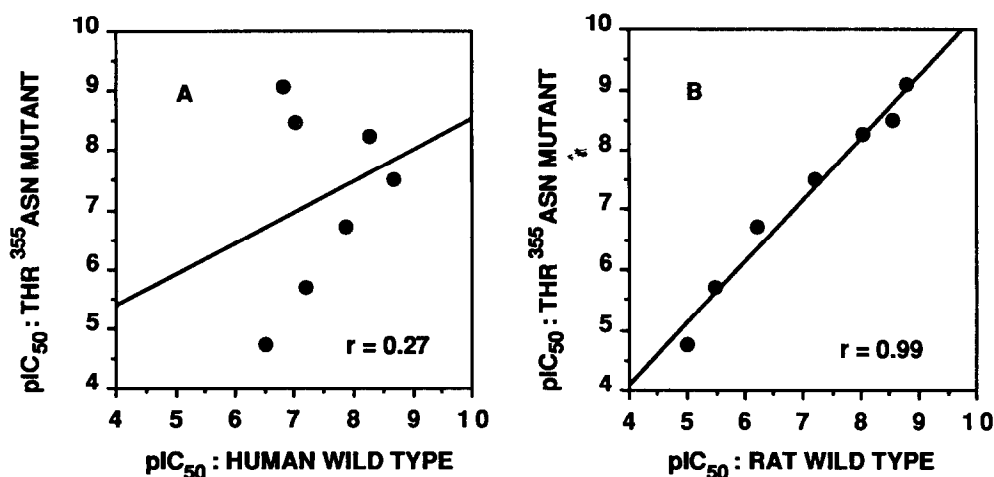


Fig. 1. Correlation between observed pIC₅₀ values of drugs competing for [³H]5-HT binding to Thr³⁵⁵Asn mutant human 5-HT_{1D} β receptors and (A): wild type human 5-HT_{1D} β receptors and (B): wild type rat 5-HT_{1D} β (5-HT_{1B}) receptors. Rat IC₅₀ values are from Ref. 3. The IC₅₀ for 8-OH-DPAT at the rat receptor was determined to be >10 μ M, and so this value was used for plotting.

The mutation-induced changes in drug affinity were emphasized by the poor correlation of the pIC₅₀ values for the mutant receptor with the values for the wild type receptor (Fig. 1A). In contrast, there was a high correlation of the pIC₅₀ values of the mutant human receptor with those of the rat 5-HT_{1B} receptor (Fig. 1B). These results confirm that the residue occupying the mutated position is a key determinant of serotonin receptor pharmacological selectivity, but they do not rule out the possibility that other amino acid residue differences may also contribute to the inter-species pharmacological difference observed in a more minor way. Data such as these in combination with molecular modeling may eventually shed light on the physical nature of ligand/receptor binding.

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REFERENCES

1. Hamblin MW and Metcalf MA, Primary structure and functional characterization of a human 5-HT_{1D}-type serotonin receptor. *Mol Pharmacol* 40: 143-148, 1991.
2. Hamblin MW, Metcalf MA, McGuffin RW and Karpells S, Molecular cloning and functional characterization of a human 5-HT_{1B} serotonin receptor: A homologue of the rat 5-HT_{1B} receptor with 5-HT_{1D} - like pharmacological specificity. *Biochem Biophys Res Commun* 184: 752-759, 1992.

3. Hamblin MW, McGuffin RW, Metcalf MA, Dorsa DM and Merchant KM, Distinct 5-HT_{1B} and 5-HT_{1D} receptors in rat: Structural and pharmacological comparison of the two cloned receptors. *Mol Cell Neurosci*, in press.
4. Jin H, Oksenberg D, Ashkenazi A, Peroutka SJ, Duncan AMV, Rozmahel R, Yang Y, Mengod G, Palacios JM and O'Dowd BF, Characterization of the human 5-hydroxytryptamine_{1B} receptor. *J Biol Chem* **267**: 5735-5738, 1992.
5. Levy FO, Gudermann T, Perez-Reyes E, Birnbaumer M, Kaumann AJ and Birnbaumer L, Molecular cloning of a human serotonin receptor (S12) with a pharmacological profile resembling that of the 5-HT_{1D} subtype. *J Biol Chem* **267**: 7553-7562, 1992.
6. Weinshank RL, Zgombick JM, Macchi MJ, Branchek TA and Hartig PR, Human serotonin 1D receptor is encoded by a subfamily of two distinct genes: 5-HT_{1D α} and 5-HT_{1D β} . *Proc Natl Acad Sci USA* **89**: 3630-3634, 1992.
7. Demchyshyn L, Sunahara RK, Miller K, Teitler M, Hoffman BJ, Kennedy JL, Seeman P, Van Tol HHM and Niznik HB, A human serotonin 1D receptor variant (5HT1D β) encoded by an intronless gene on chromosome 6. *Proc Natl Acad Sci USA* **89**: 5522-5526, 1992.
8. Adham N, Romanienko P, Hartig P, Weinshank RL and Branchek T, The rat 5-hydroxytryptamine_{1B} receptor is the species homologue of the human 5-hydroxytryptamine_{1D β} receptor. *Mol Pharmacol* **41**: 1-7, 1992.
9. Voigt MM, Laurie DJ, Seeburg PH and Bach A, Molecular cloning and characterization of a rat brain cDNA encoding a 5-hydroxytryptamine_{1B} receptor. *EMBO J* **10**: 4017-4023, 1991.
10. Maroteaux L, Saudou F, Amlaiky N, Boschert U, Plassat JL and Hen R, Mouse 5HT1B serotonin receptor: cloning, functional expression, and localization in motor control centers. *Proc Natl Acad Sci USA* **89**: 3020-3024, 1992.
11. Hoyer D and Middlemiss DN, Species differences in the pharmacology of terminal 5-HT autoreceptors in mammalian brain. *Trends Pharmacol Sci* **10**: 130-132, 1989.
12. Kobilka BK, Kobilka TS, Daniel K, Regan JW, Caron MG and Lefkowitz RJ, Chimeric α_2 - β_2 -adrenergic receptors: delineation of domains involved in effector coupling and ligand binding specificity. *Science* **240**: 1310-1316, 1988.
13. Suryanarayana S, Daunt DA, Von Zastrow M and Kobilka BK, A point mutation in the seventh hydrophobic domain of the α_2 adrenergic receptor increases its affinity for a family of β receptor antagonists. *J Biol Chem* **266**: 15488-15492, 1991.
14. Guan X-M, Peroutka SJ and Kobilka BK, Identification of a single amino acid residue responsible for the binding of a class of β -adrenergic receptor antagonists to 5-hydroxytryptamine_{1A} receptors. *Mol Pharmacol* **41**: 695-698, 1992.
15. Ito W, Ishiguro H and Kurosawa Y, A general method for introducing a series of mutations into cloned DNA using the polymerase chain reaction. *Gene* **102**: 67-70, 1991.
16. Cullen BR, Use of eukaryotic expression technology in the functional analysis of cloned genes. In: *Guide to Molecular Cloning Techniques* (Eds. Berger SL and Kimmel AR), pp. 684-704. Academic Press, San Diego, 1987.
17. Parker RB and Waud DR, Pharmacological estimation of drug-receptor dissociation constants. Statistical evaluation. I. Agonists. *J Pharmacol Exp Ther* **177**: 1-12, 1971.
18. Waeber C, Schoeffer P, Palacios JM and Hoyer D, Molecular pharmacology of 5-HT_{1D} sites. *Naunyn-Schmiedeberg's Arch Pharmacol* **337**: 595-601, 1988.
19. Herrick-Davis K, Titler M, Leonhardt S, Struble R and Price D, Serotonin 5-HT_{1D} receptors in human prefrontal cortex and caudate: Interaction with a GTP binding protein. *J Neurochem* **51**: 1906-1912, 1988.
20. Peroutka SJ, Switzer JA and Hamik A, Identification of 5-hydroxytryptamine_{1D} binding sites in human brain membranes. *Synapse* **3**: 61-66, 1989.
21. Hoyer D, Engel G and Kalkman HO, Molecular pharmacology of 5-HT₁ and 5-HT₂ recognition sites in rat and pig brain membranes: Radioligand binding studies with [³H]5-HT, [³H]8-OH-DPAT, (-)[¹²⁵I]iodocyanopindolol, [³H]mesulergine and [³H]ketanserin. *Eur J Pharmacol* **118**: 13-23, 1985.